

1 **SUPPORTING INFORMATION**

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4 **Transformation of pyrene in aqueous chlorination in the presence and absence of bromide**  
5 **ion: Kinetics, Products and Their Aryl Hydrocarbon Receptor-Mediated Activities**  
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1 ***Kinetics Experiments.*** Tested aqueous solutions (200mL) were buffered using phosphate salts  
2 for 5.5-9.0 pH range. For pH < 5.5 and pH > 9, pH values were preadjusted with H<sub>2</sub>SO<sub>4</sub> and  
3 NaOH, respectively. The initial pyrene concentration was 0.5 μM, at least 35 μM of total  
4 chlorine was added. Chlorine variation was less than 5% under these conditions. The chlorine  
5 concentration was thus assumed to be constant during the kinetics experiments.

6 Kinetic runs were initiated by injecting, under rapid mixing, an aliquot of sodium  
7 hypochlorite solution. At different reaction times, 3 ml of solution was rapidly transferred into a  
8 vial containing 100 μL of sodium thiosulfate solution (1 M) to quench the residual chlorine and  
9 stop the reaction. Samples were then analyzed using HPLC to determine the remaining pyrene  
10 concentration. When the pyrene disappeared, the kinetic experiments were pursued until at least  
11 50% pyrene consumption was achieved.

12 Each sample was analyzed by HPLC with a reversed-phase C18 column (Zorbax-ODS, 4.6  
13 mm I.D. × 250 mm in length, 5 μm in particle diameter, Alliance and Agilent, USA) at 25°C.  
14 The initial mobile phase composition was acetonitrile /water (70/30 v/v), which was increased  
15 linearly to 100% acetonitrile in 10 min., and then held for 15 min. The flow rate, UV-detection  
16 wavelength, and sample injection volume were 1.5 mL/min, 333 nm, and 50 μL, respectively.

17 ***Identification and analysis of by-products.*** The experiments were carried out in a glass reactor  
18 which was placed in a water bath to maintain the reaction temperature at 25°C. Synthetic raw  
19 water was prepared by dissolving 0.25 mg of a standard pyrene into 2.5 L Milli-Q pure water of  
20 which the pH was adjusted to 7.22 by adding phosphate buffer. A 500-mL sample was removed  
21 for determination of the aryl hydrocarbon receptor mediated activity before NaOCl was added to  
22 the remaining solution. Samples (500 mL) were taken out at different chlorination time after  
23 NaOCl was added to the remaining solution ([HOCl]<sub>T</sub> = 35 μM). After decomposition of the  
24 residual HOCl by the addition of Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (1 M) and after the pH was adjusted to 2-3, the  
25 samples were concentrated by solid phase extraction (SPE, Waters Sep-Pak C18, Waters, USA).  
26 The cartridge was conditioned with 5ml dichloromethane, 5ml methanol and 5ml water. Samples

1 were passed through the cartridge at a flow rate of 10-15 mL/min. Additional water (10mL) was  
2 applied to wash the wall of the cartridge. The residual water was removed by passing a gentle  
3 nitrogen stream through the cartridge for about 30 min. 10mL dichloromethane was percolated at  
4 4-5mL/min through the sorbent bed to mobilize pyrene and its chlorinated products. Of the  
5 10mL eluant, 8mL was dried under a gentle nitrogen stream, and redissolved in 0.2mL hexane  
6 for GC-MS analysis. The rest of the eluant (2mL) was dried and redissolved in 0.02mL DMSO  
7 to assess the AhR-mediated activity using the yeast two-hybrid assay. The products in an  
8 aqueous chlorinated solution of pyrene were analyzed by GC-MS (Hewlett-Packard  
9 (5890-5971)), and identified by NMR.

10 The column was a HP-5MS (60m × 0.32mm × 0.25μm, J & W Scientific, USA). Splitless  
11 injections were employed with 5-psi head pressure and 2 mL/min He carrier flow rate. The  
12 temperature was increased from 70°C to 210°C at the rate of 15°C/min, then raised to 300°C at  
13 the rate of 8°C /min (and held for 10 min) for analysis of the aqueous chlorinated samples  
14 without the addition of bromide ion. For the analysis of samples with the addition of bromide  
15 ion, the temperature program was 70°C to 300°C at the rate of 8 °C/min (held for 10 min). The  
16 mass range scanned was 50 to 550 amu with a scan time of 1.6 sec.

17 The <sup>1</sup>H NMR and H-H COSY spectra were measured on a Bruker ARX400 (<sup>1</sup>H, 400 MHz,  
18 Switzerland) instrument. Deuteriochloroform was used as the solvent. The chemical shifts (δ  
19 values) are given in ppm downfield from tetramethylsilane.

20 ***Synthesis of 1,8-dibromopyrene and 1,6-dibromopyrene.*** Di-Br-pyrene standards were obtained  
21 by the reaction of 1-bromopyrene with sodium hypochlorite in the presence of potassium  
22 bromide in acidic 50% methanol solution. This product was suggested to be a mixture of two  
23 isomers by GC/MS analysis. Separation of two isomers was successful by following fractional  
24 recrystallization. The mixture was dissolved in hot hexane and allowed to stand at room  
25 temperature to deposit colorless crystals, which were collected on a suction filter. These crystals  
26 were 1,8-dibromopyrene, mp 206-208 °C. The residual solid obtained by concentration of the

1 filtrate was recrystallized twice from hexane to give pure crystals of 1,6-dibromopyrene, mp  
2 224-227 °C.

3 ***Yeast Assay for AhR-mediated Activity of Products.*** The yeast strain YCM3 with human AhR,  
4 Arnt and the LacZ reporter plasmid, pTXRE5-Z, was used to test the AhR-mediated activity. The  
5 human AhR and Arnt genes are integrated into chromosome III. AhR and Arnt are expressed  
6 from the galactose-regulated GAL 1,10 promoter. Transcriptional activation mediated by the  
7 AhR/Arnt heterodimer is assessed by  $\beta$ -galactosidase activity. Expression of the LacZ reporter  
8 plasmid, TXRE5-Z, is directed by the AhR-Arnt complex binding to five response elements in  
9 the promoter region. The yeast cells were preincubated at 30°C for 22 hours in 5 mL medium  
10 (6.7 g/L Difco yeast nitrogen base without amino acids, 0.2% glucose, 117.6 mg/L L-Leucine,  
11 300 mg/L L-isoleucine, 1500 mg/L L-valine, 200 mg/L L-adenine hemisulfate salt, 200 mg/L  
12 L-arginine HCl, 200 mg/L L-histidine HCl monohydrate, 300 mg/L L-lysine HCl, 200mg/L  
13 L-methionine, 500mg/L L-phenylalanine, 200 mg/L L-threonine, 300 mg/L L-tyrosine, 200mg/L  
14 L-uracil (Sigma, USA)). 50  $\mu$ L of overnight culture and 2.5  $\mu$ L of DMSO solution diluted to the  
15 desired concentrations were then added to 200  $\mu$ L of fresh medium (2% galactose) in a glass tube  
16 (10 mm  $\times$  50 mm), respectively. After yeasts were cultured for 8 h at 30°C, 150  $\mu$ L of the above  
17 culture was fractionated, and its absorbance at 595 nm was detected. The residual culture (100  
18  $\mu$ L) was centrifuged at 4 °C (15000 rpm) for 5 min, and the collected cells were resuspended in  
19 200  $\mu$ L of Z buffer (0.1 M sodium phosphate (pH = 7.0), 10 mM KCl, 1 mM MgSO<sub>4</sub>) containing  
20 1mg/mL Zymolyase 20T (Seikagaku, Tokyo), and incubated for 20 min at 30°C. The enzymatic  
21 reaction was started by the addition of 40  $\mu$ L of 4 mg/mL 2-nitrophenyl- $\beta$ -D-galactopyranoside  
22 (ONPG, Tokyo Kasei, Tokyo, Japan), and incubated for 20 min at 30°C. Then the enzymatic  
23 reaction was stopped by adding 1 M Na<sub>2</sub>CO<sub>3</sub> (100  $\mu$ L). After the above solution was centrifuged,  
24 150- $\mu$ L aliquots were placed into 96 wells of a microplate. Absorbances at 415 and 570nm were  
25 read on a microplate reader (Bio RAD 550, USA) to estimate the AhR-mediated activity, and the  
26  $\beta$ -galactosidase activity (U) was calculated according to Equation (1):

$$U = \frac{1000 \times (OD_{415} - 1.75 \times OD_{570})}{v \times t \times OD_{595}} \quad (1)$$

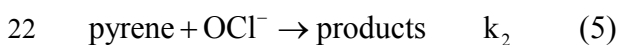
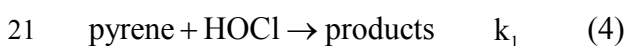
where  $t$  represents the reaction time (min),  $v$  is the volume of the culture used in the assay (mL),  $OD_{595}$  is the cell density at the start of the assay,  $OD_{415}$  is the absorbance by *o*-nitrophenol at the end of the reaction, and  $OD_{570}$  is the light scattering at the end of the reaction. In this assay,  $\beta$ -naphthoflavone (Chemservice, Chester, England) was used as positive control.

**Kinetics of chlorination of pyrene.** The kinetics of pyrene chlorination were investigated under conditions for pseudo-first-order kinetics ( $[HOCl]_0 > 20 \times [pyrene]_0$ ). The initial concentration of pyrene was 0.5  $\mu$ M and at least 35  $\mu$ M of chlorine was added. Chlorine variation was less than 5% under these conditions. The concentration of chlorine was assumed to be constant during the reaction time. Figure 1 shows the plot of  $\ln([pyrene]_t/[pyrene]_0)$  as a function of reaction time at pH = 5.01, 6.57 and 7.22. The results show that the kinetic experiments can be well described by a pseudo-first-order kinetics as demonstrated by the linear plots ( $r^2 > 0.97$ ). The reaction order with respect to HOCl was examined at pH 7.22 by varying its concentration. The insert in Figure 1 shows that the pseudo-first-order rate constant,  $k_{obs}$ , is linearly correlated to the chlorine dose. Therefore, the rate of pyrene disappearance is first order to total concentration of chlorine ( $[HOCl]_T = [HOCl] + [OCl^-]$ ) and to the pyrene concentration ( $[pyrene]$ ):

$$v = -\frac{d[pyrene]}{dt} = k_{app}^{Cl} [pyrene] [HOCl]_T \quad (2)$$

with  $k_{app}^{Cl} = k_{obs} / [HOCl]_T$  is a second-order kinetic constant.

The pyrene chlorination kinetics can be expressed as follows:



and the rate expression for the above reaction is

$$v = -\frac{d[\text{pyrene}]}{dt} = k_1[\text{HOCl}][\text{pyrene}] + k_2[\text{OCl}^-][\text{pyrene}] \quad (6)$$

By replacing  $[\text{HOCl}]$  and  $[\text{OCl}^-]$  as ratios of  $[\text{HOCl}]_T$ , the rate of pyrene disappearance is

$$v = -\frac{d[\text{pyrene}]}{dt} = k_1\alpha_1^{\text{Cl}}[\text{HOCl}]_T[\text{pyrene}] + k_2\alpha_2^{\text{Cl}}[\text{HOCl}]_T[\text{pyrene}] \quad (7)$$

where  $\alpha_i^{\text{Cl}}$  is the ionization fraction of hypochlorous acid species, with  $i = 1$  or  $2$ , for  $\text{HOCl}$  and  $\text{OCl}^-$ , respectively.

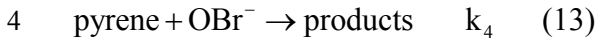
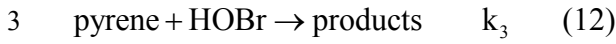
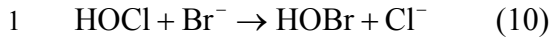
Therefore, by combining Eqs (2) and (7),  $k_{\text{app}}^{\text{Cl}}$  is a function of pH

$$k_{\text{app}}^{\text{Cl}} = k_1\alpha_1^{\text{Cl}} + k_2\alpha_2^{\text{Cl}} = k_1 \frac{[\text{H}^+]}{K_{\text{aCl}} + [\text{H}^+]} + k_2 \frac{K_{\text{aCl}}}{K_{\text{aCl}} + [\text{H}^+]} \quad (8)$$

In order to investigate the effect of bromide ion on the kinetics of reaction, 5  $\mu\text{M}$  bromide ions were added in water before chlorination reaction. Hypobromous acid ( $\text{HOBr}$ ) is formed because of reaction with chlorine ( $\text{HOCl} + \text{Br}^- \rightarrow \text{HOBr} + \text{Cl}^-$ ). This reaction is fast. So equal molar of hypobromous acid (equal to the concentration of bromide ions added) was formed. The experimental procedure was as the same as described above. Figure 2 shows the plot of  $\ln([\text{pyrene}]_t/[\text{pyrene}]_0)$  as a function of reaction time at  $\text{pH} = 6.80, 7.82$  and  $8.27$ . The results show that the kinetic experiments can be well described by a pseudo-first-order kinetics as demonstrated by the linear plots ( $r^2 > 0.96$ ). The reaction order with respect to bromide ions was examined at  $\text{pH} 8.27$  by varying its concentration. The insert in Figure 2 shows that the pseudo-first-order rate constant,  $k_{\text{obs}}$ , is linearly correlated to the concentration of total concentration of hypobromous acid and the existence of  $\text{HOCl}$  do not significantly contribute to the overall reaction rate. Therefore, the rate of pyrene disappearance is first order to concentration of hypobromous acid ( $[\text{HOBr}]_T = [\text{HOBr}] + [\text{OBr}^-]$ ) and to the pyrene concentration ( $[\text{pyrene}]$ ):

$$v = -\frac{d[\text{pyrene}]}{dt} = k_{\text{obs}}[\text{pyrene}] = k_{\text{app}}^{\text{Br}}[\text{HOBr}]_T[\text{pyrene}] \quad (9)$$

The reactions in the presence of bromide ions can be expressed as follows:



5 The rate expression can be written as follows:

6  $v = k_3[\text{pyrene}][\text{HOBr}] + k_4[\text{pyrene}][\text{OBr}^-]$  (14)

7 By replacing  $[\text{HOBr}]$  and  $[\text{OBr}^-]$  as ratios of  $[\text{HOBr}]_{\text{T}}$ , the rate of pyrene disappearance is

8  $v = -\frac{d[\text{pyrene}]}{dt} = k_3\alpha_1^{\text{Br}}[\text{HOBr}]_{\text{T}}[\text{pyrene}] + k_4\alpha_2^{\text{Br}}[\text{HOBr}]_{\text{T}}[\text{pyrene}]$  (15)

9 where  $\alpha_i^{\text{Br}}$  is the ionization fraction of hypochlorous acid species, with  $i = 1$  or  $2$ , for HOBr and

10  $\text{OBr}^-$ , respectively.

11 Therefore, by combining Eqs (9) and (15),  $k_{\text{app}}^{\text{Br}}$  is a function of pH

12  $k_{\text{app}}^{\text{Br}} = k_3\alpha_1^{\text{Br}} + k_4\alpha_2^{\text{Br}} = k_3 \frac{[\text{H}^+]}{K_{a_{\text{Br}}} + [\text{H}^+]} + k_4 \frac{K_{a_{\text{Br}}}}{K_{a_{\text{Br}}} + [\text{H}^+]}$  (16)

13 **Calculation of Model Selection Criterion (MSC).** We employed MSC as an index to estimate  
 14 the goodness of fit for the regressions in Figure 1. The MSC is an extension of Akaike  
 15 information criterion (AIC), which is derived from the maximum likelihood function ( $I$ ). The  
 16 MSC is estimated as following equation:

17 
$$\ln \left[ \frac{\sum_{i=1}^n W_i (x_i - \bar{x}_i)^2}{\sum_{i=1}^n W_i (x_i - \tilde{x}_i)^2} \right] - \frac{2p}{n}$$

18 where  $x_i$  represents the  $i$ -th observed data,  $\tilde{x}_i$  is the  $i$ -th predicted value,  $\bar{x}_i$  is the mean  
 19 observed value,  $n$  is the number of data,  $p$  is the number of parameters, and  $W_i$  is the weighting  
 20 of data. The value of  $W_i$  depends on the uncertainty of data. In general, the  $W_i$  should be 1. The  
 21 higher the MSC, the closer the model explains the observed values. The theoretical models that  
 22 produce MSC values larger than 3 are regarded to exhibit an acceptable fit to data, whereas

1 exceptionally good fit ( $MSC > 6$ ) should be taken as suspects, and the value less than 2 was  
2 regarded unacceptable. As a result, the MSCs for the regressions (Figure 1) in the absence and  
3 presence of bromide ions were 3.10 and 3.85, respectively.

4

#### 5 **Literature Cited**

6 (1) Micromath Scientific Software. *RSTRIP Polyexponential Curve Stripping/Least Squares*  
7 *Parameter Estimation*. Micromath Scientific Software: Salt Lake City, UT, USA. 1989.

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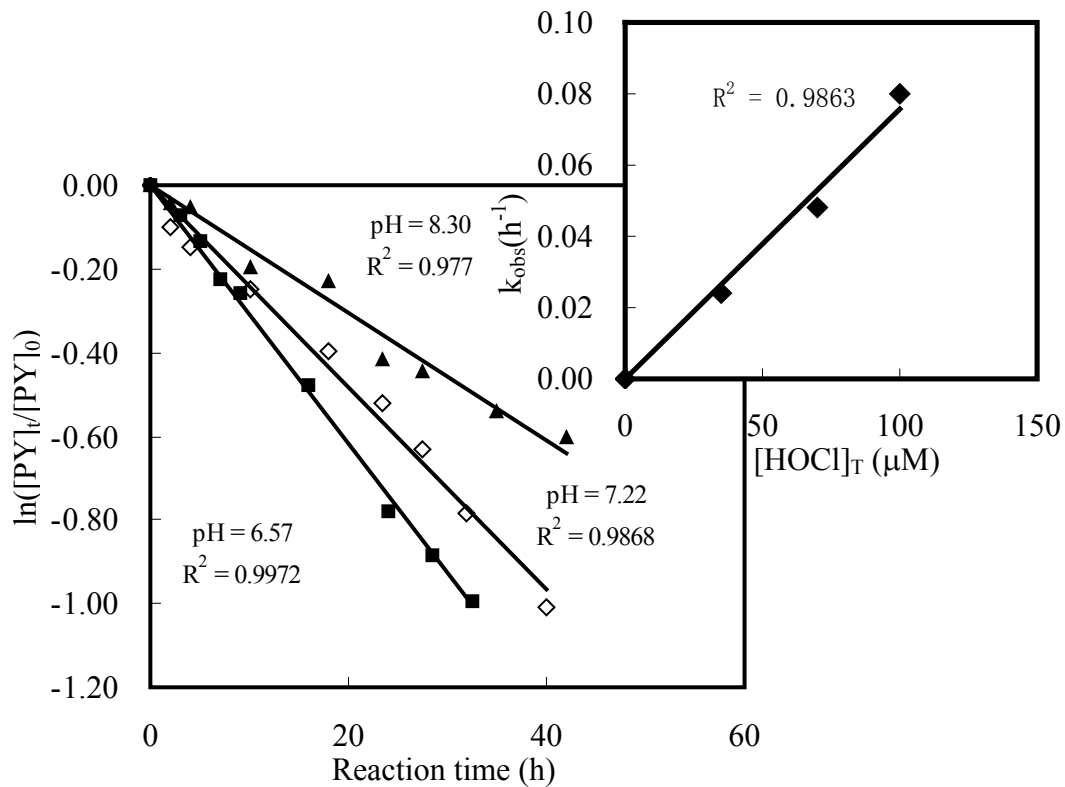
1 **SUPPORTING INFORMATION Table 1.** GC/MS mass spectra of products in aqueous chlorinated solution  
2 of pyrene.

	Retention time	Molecular ion	m/z (relative abundance)
pyrene	17.287	202	202(100), 203(17), 200(15), 201(14), 101(99)
1-Cl-pyrene	19.530	236	236(100), 238(32), 200(30), 100(21), 235(18)
1-Br-pyrene	20.714	280	280(100), 282(98), 201(68), 200(65), 100(51)
di-Cl-pyrene	21.408	270	270(100), 272(66), 200(45), 100(20), 274(10)
	21.728	270	
Br-Cl-pyrene	22.809	316	314(100), 316(75), 200(61), 100(35), 101(20)
	23.169	316	
di-Br-pyrene	24.369*	360	360(100), 362(52), 358(49), 281(25), 200(69)
	24.769*	360	

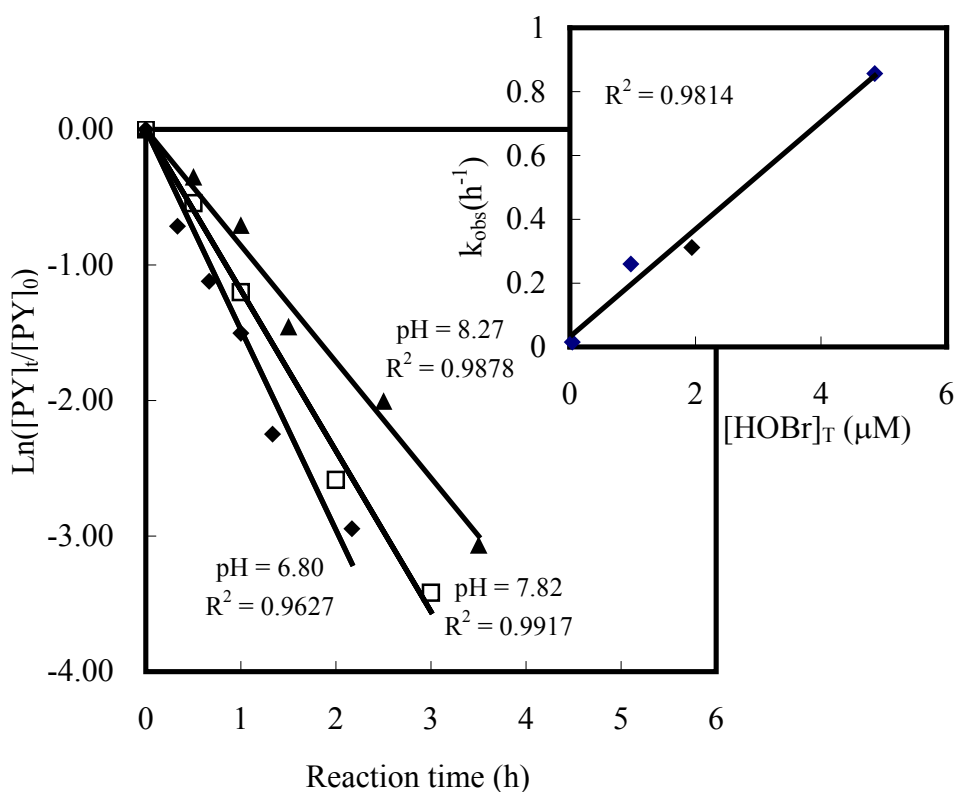
3 \* Retention time in Figure 3.

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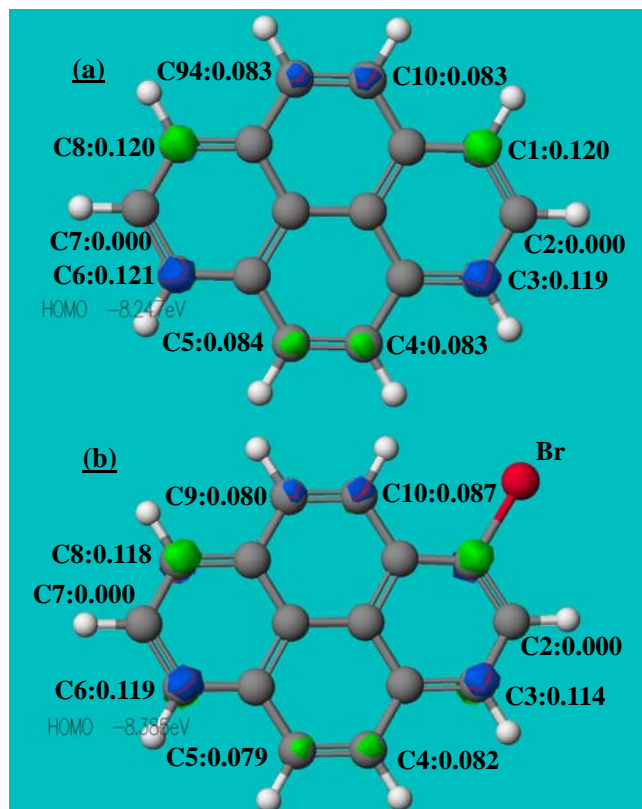
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**SUPPORTING INFORMATION Figure 1.** Pseudo-first-order kinetic plot of pyrene chlorination at  $20 \pm 2$  °C,  $[HOCl]_T = 35 \mu M$  and three pH levels. Symbols represent measured data, and the straight line is the linear regression. (Insert: pH = 7.22,  $20 \pm 2$  °C, and various  $[HOCl]_T$ )



**SUPPORTING INFORMATION Figure 2.** Pseudo-first-order kinetic plot of pyrene chlorination at  $20 \pm 2$  °C,  $[HOCl]_T = 35$   $\mu M$  and three pH levels with addition of bromide ions (5  $\mu M$ ). Symbols represent measured data, and the straight line is the linear regression. (Insert: pH = 9.20,  $20 \pm 2$  °C,  $[HOCl]_T = 35$   $\mu M$  and various  $[Br^-]$ )



**SUPPORTING INFORMATION** Figure 3. Atom HOMO (highest occupied molecular orbital) density of pyrene and 1-Br-pyrene. (a) pyrene; (b)1-Br-pyrene. Atom HOMO density were calculated by MOPAC Ver.6 (CACHe Scientific Inc., Oxford)